

Tipping the metabolic scales towards increased longevity in mammals

Celine E. Riera and Andrew Dillin

A hallmark of ageing is dysfunction in nutrient signalling pathways that regulate glucose homeostasis, negatively affecting whole-body energy metabolism and ultimately increasing the organism's susceptibility to disease. Maintenance of insulin sensitivity depends on functional mitochondrial networks, but is compromised by alterations in mitochondrial energy metabolism during ageing. Here we discuss metabolic paradigms that influence mammalian longevity, and highlight recent advances in identifying fundamental signalling pathways that influence metabolic health and ageing through mitochondrial perturbations.

Life expectancy has been strikingly prolonged in developed countries as a positive outcome of medical progress and modern lifestyle. Nonetheless, with old age comes a wide range of age-associated diseases including neurodegenerative diseases, cardiovascular and metabolic disorders, and higher susceptibility to cancer.

Although the clinical manifestations of ageing are well understood, the genetic or environmental causes driving age-associated decline in healthspan remain difficult to isolate. By identifying genetic perturbations that can extend lifespan as well as retard the onset of physiological decline in invertebrate model organisms, ageing research has started to provide therapeutic strategies to prolong both healthspan and lifespan¹. Research over the last decade using the multitude of spatial and temporal genetic tools available in mice has begun to identify points of convergence between mammals and invertebrates. In comparison to humans, mice are relatively short-lived mammals that suffer from clinically relevant pathologies including metabolic syndrome, osteoporosis, cancer, sarcopenia, cardiovascular dysfunction and neurodegeneration^{2,3}. Molecular and cellular pathways have been identified that not only increase longevity of mammalian model organisms but also generally promote their healthspan by delaying the onset of metabolic decline. This delay is tightly coupled to an age-dependent deterioration of insulin metabolism and subsequent disruption of nutrient uptake by body tissues. Mitochondria play a key role in the utilization of nutrient substrates for energy production. Alterations in mitochondrial biogenesis, dynamics and integrity render an organism unable to maintain metabolic homeostasis or properly respond to metabolic demand observed during ageing.

In this Review, we provide an overview of the metabolic signalling pathways that modulate mammalian lifespan and discuss their potential as therapeutic targets to treat age-associated loss in glucose homeostasis and healthspan. Finally, we present key mitochondrial

pathways that have been implicated in age-dependent malfunction of metabolic homeostasis.

Insulin action regulates metabolic health during ageing

The pathologies encountered in mouse longevity studies revealed a dominant role for glucose homeostasis in regulating metabolic health during ageing. In humans, ageing is also linked to increased visceral fat mass, loss in lean mass and a deterioration in insulin sensitivity resulting in glucose intolerance — all factors that favour the occurrence of metabolic syndrome and cardiovascular diseases^{4,5}. Similarly, in mice, signs of physiological decline are characterized by an increase in fat mass and the progressive development of glucose intolerance, reflecting a similar impairment of insulin sensitivity^{6,7}. However, mild insulin resistance has been observed in a variety of long-lived mice and is therefore not necessarily an indicator of poor health or shortened lifespan when paired with improved glucose tolerance^{7–10}. Over the last decade, elevated energy expenditure has emerged as a putative longevity biomarker, as it has been associated with lifespan extension¹¹. In old animals, a decrease in energy expenditure is associated with reduced activity and endurance linked to decreased oxygen consumption and lipid accumulation in non-adipose tissue. The respiratory exchange ratio (RER), obtained by indirect calorimetry, compares the volume of carbon dioxide an organism produces to the volume of oxygen consumed over a given time and varies inversely with lipid oxidation. In young and healthy mice, the RER displays a youthful circadian shift from night to day, reflecting the daily transition from carbohydrate to lipid metabolism. Old mice, however, develop a substrate preference towards lipids, losing the capacity to switch between fuel sources (Fig. 1). This is in accordance with a metabolic flexibility theory of ageing, which predicts that longevity depends on metabolic health, and lifespan extension is achieved by maintaining a youthful RER, thus

Celine E. Riera and Andrew Dillin are at the Department of Molecular and Cell Biology, University of California, Berkeley, California 94720, USA and Howard Hughes Medical Institute, Chevy Chase, Maryland 20815, USA.
e-mail: dillin@berkeley.edu

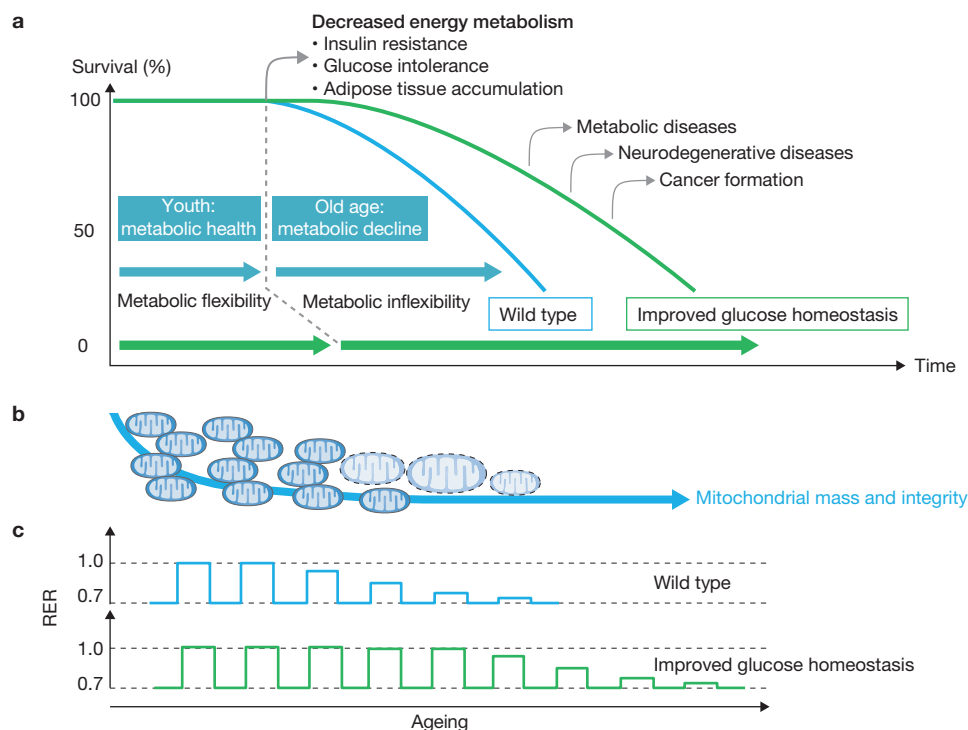


Figure 1 Metabolic flexibility controls healthspan and lifespan. **(a)** A schematic showing survival curves of a wild-type mouse (blue curve) compared to a long-lived genetic model of improved glucose homeostasis (green curve). The onset of metabolic decline precedes the appearance of morbid pathologies associated with insulin resistance and nutrient uptake. **(b)** Throughout lifespan, mitochondrial biogenesis, function and dynamics decline. Accumulation of oversized abnormal mitochondria which result

from impaired degradation of dysfunctional mitochondria is observed after the onset of metabolic inflexibility. **(c)** Metabolic flexibility correlates with a healthy respiratory exchange ratio (RER) resulting from the circadian shift between carbohydrate (value of 1.0) to lipid metabolism (value of 0.7). Metabolic inflexibility observed at old age correlates with a substrate preference towards lipids, reflecting an inability to adapt fuel oxidation to fuel availability.

protecting the organism against systemic damage associated with fat metabolism and storage. In contrast, the onset of metabolic inflexibility corresponds to aberrant glucose homeostasis and increased susceptibility to age-onset diseases in response to an age-associated decline in mitochondrial energy metabolism¹². Another mechanism that may contribute to the deterioration of metabolic health with age operates through the oscillations of the circadian clock. Modulating the activity of the major circadian regulators BMAL1 (also known as ARNTL; aryl hydrocarbon receptor nuclear translocator-like) and CLOCK (clock circadian regulator) results in premature ageing¹³, whereas enhanced transcriptional activity of these factors protects against age-associated decline in adapting to new circadian periods¹⁴. Most physiological, endocrine and behavioural rhythms are governed by the molecular clock, and accumulating evidence indicates that disruption of circadian rhythms is associated with cancer¹⁵. Future research is required to shed light on how the rhythmicity of nutrient homeostatic processes affects metabolic flexibility and the ageing process.

Genetic manipulation of insulin metabolism in mouse lifespan and healthspan

To date, dietary restriction is the most robust intervention to increase healthspan and lifespan in multiple organisms including rodents and primates, delaying the onset of age-related diseases^{2,16,17}. In mice, dietary restriction increases insulin sensitivity and induces a healthy transition of the RER, due to a more dramatic shift from carbohydrate oxidation

and endogenous fatty acid synthesis in the feeding phase to fatty acid oxidation during resting¹⁸. Similarly to dietary restriction, multiple genetic manipulations in mice improve metabolic flexibility and longevity. Alterations of components in the insulin and insulin-like growth factor signalling (IIS) pathway that dampen insulin signalling promote longevity associated with improved insulin sensitivity. Some of these long-lived mice include fat-specific insulin receptor knockout (FIRKO) mice, hypomorphic PI3K mice, and mice overexpressing the liver hormone FGF21 (fibroblast growth factor 21), which antagonizes growth hormone (GH) and insulin-like growth factor 1 (IGF-1)^{19–21}. Reduction of circulating IGF-1 positively correlates with increased longevity in inbred mouse strains²², but is also linked to a reduced body size and even dwarfism²³.

Although Ames (*Prop1^{df}/Prop1^{df}*) and Snell (*Pit1^{dw}/Pit1^{dw}*) dwarf mice are extremely long-lived presumably partially because of decreased IIS, their metabolic characteristics must be interpreted with caution. This is because profound deficiency in anterior pituitary function results in other hormonal changes and negatively affects fertility, metabolic fitness, adiposity, glucose tolerance and insulin secretion, despite considerable life extension^{24,25}. In humans, hypothyroidism is also characterized by diminution of the acute insulin response, resulting in impaired glucose tolerance²⁶, but the effects of this condition on human longevity are unclear. Thus, the lifespan extension of reduced IIS models must be uncoupled from their growth delay or hormonal imbalances to isolate clinically relevant therapeutic targets.

Deletion of TRPV1 (transient receptor potential vanilloid 1) pain receptors in peripheral nerves innervating pancreatic β cells extends mouse longevity by improving metabolic flexibility and in particular elevating prandial insulin secretion with age, without the corresponding growth delay or altered body weight observed in decreased IIS models⁷. In humans, metabolic syndrome is linked to dementia and increased incidence of multiple cancers^{27,28}. Strong correlations between insulin sensitivity and cancer formation have also been found in the mouse, and reduced cancer incidence is observed in mice with improved energy expenditure at old age^{7,29}. Mice carrying a TRPV1 mutation have both reduced cancer incidence and delayed onset of cognitive decline with age. Other genetic manipulations that reduce IIS meet these criteria to some extent. Long-lived Ames dwarf mice and GHR-BP (growth-hormone receptor binding protein) knockout mice display a lower incidence and delayed onset of certain cancers and tumours, increased insulin sensitivity and improved cognitive function at old age^{30–32}. Additionally, prolonged rapamycin treatment delays cancer formation in aged mice and extends lifespan through inhibition of mTOR (mammalian target of rapamycin), a major cellular nutrient sensing pathway that regulates cell growth^{33,34} and integrates insulin signalling and cell stress signals³⁵. The method by which rapamycin regulates the rate of ageing is complex, because short-term treatment results in immunodeficiency and favours glucose intolerance and insulin resistance^{36,37}. However, prolonged treatment results in improved metabolic profiles, increased oxygen consumption and ketogenesis, and markedly enhanced insulin sensitivity³⁸. Robust genetic inhibition of downstream mTOR complex 1 (mTORC1) components, including deletion of ribosomal protein S6K1 (S6 kinase 1) and enhanced autophagy, lead to increased longevity with improved glucose tolerance and insulin sensitivity at mid-age^{39,40}. However, despite the consensus that dampening mTORC1 activity extends lifespan, several studies report that this beneficial effect is achieved mostly by slowing cellular growth and cancer incidence, without impacting insulin metabolism^{36,41}. These reports suggest that disruption of the other major mTOR complex, mTORC2, mediates aspects of the impaired insulin metabolism associated with rapamycin treatment, in particular the inability to suppress hepatic gluconeogenesis following insulin release. It is therefore unclear how prolonged rapamycin exposure promotes metabolic health, despite the recent report that insulin sensitivity amelioration following rapamycin treatment might rely on inhibitory effects of both mTOR complexes³⁸. Better resolution of the mechanism through which rapamycin positively modulates glucose homeostasis is required to design drugs that can recapitulate the longevity extension obtained by rapamycin treatment without its detrimental side effects.

Mitochondria at the core of metabolic flexibility during ageing

Even though cellular energy can be rapidly generated through anaerobic glycolysis, it mostly originates from aerobic oxidation of carbohydrates and fatty acids in the mitochondria, highlighting this organelle's critical role in regulating global metabolic homeostasis. Not only is insulin secretion from pancreatic β cells mitochondria-dependent, but insulin signalling also relies on mitochondria to metabolize glucose in energy-consuming cells. Mitochondrial activity regulates gluconeogenesis and triglyceride synthesis in the liver and lipolysis in adipose tissue, as well as many other vital processes⁴². Accordingly, mitochondrial dysfunction or damage can greatly perturb metabolic flexibility

and insulin sensitivity, impacting metabolic human diseases and ageing^{12,43,44}. During the ageing process, mitochondrial biogenesis, function and dynamics decline (Fig. 2)^{45,46}. This is particularly obvious in skeletal muscle — a highly metabolically active tissue extremely rich in mitochondria — which undergoes sarcopenia, a critical loss in mitochondrial volume density and decline in physical function with age⁴⁷. Accumulation of lipids in skeletal muscle, liver and adipose tissue is indicative of decreased metabolic flexibility with ageing and therefore demonstrates a global reduction in oxidative phosphorylation capacity and mitochondrial biogenesis.

Molecular targets that can restore mitochondrial function and metabolic homeostasis at old age have emerged over the past decade. A key regulator of mitochondrial biogenesis and energy metabolism is the transcriptional coactivator PGC1 α (proliferator-activated receptor γ coactivator α)^{48,49}. Decreased PGC1 α expression has been implicated in muscle insulin resistance in humans and diabetic mouse models^{50,51}. Increasing PGC1 α content in mouse skeletal muscle preserves oxidative phosphorylation, preventing muscle wasting and improving metabolic fitness by inhibiting insulin resistance and fat accumulation with age⁵². Mechanistically, PGC1 α acts by regulating the activity of several transcription factors — including the nuclear respiratory factors NRF1 and NRF2 and mitochondrial transcription factor A (TFAM) — that in turn directly or indirectly induce genes related to mitochondrial biogenesis and respiration^{53,54}. AMP-activated protein kinase (AMPK)-mediated phosphorylation activates PGC1 α in conditions of low cellular energy, which corresponds to a high AMP/ATP ratio, therefore inhibiting anabolic processes consuming energy and activating catabolic pathways producing energy, such as fatty acid oxidation and respiration^{55,56}. Strategies to directly manipulate AMPK to improve mitochondrial function in skeletal muscle have proven successful and offer important avenues to treat age-associated metabolic decline. Pharmacological AMPK activation by AICAR (5-Aminoimidazole-4-carboxamide ribonucleotide) treatment enhances exercise endurance⁵⁷ and increases mitochondrial biogenesis and respiration in mouse models of cytochrome *c* oxidase (COX) deficiency, rescuing the limited exercise capacity of these mice⁵⁸. Importantly, activating AMPK by metformin supplementation is also linked to lifespan extension⁵⁹, although metformin action on lifespan might depend on other targets such as mTOR (ref. 60). Under low energy conditions, AMPK also increases the ratio of nicotinamide adenine dinucleotide NAD⁺/NADH, a metabolic signal that triggers the NAD-dependent deacetylase SIRT1 (sirtuin 1) and is required for AMPK-dependent PGC1 α activation⁶¹. AMPK and SIRT1 can autoregulate each other, a mechanism that has been postulated to improve the cellular response to low-energy states⁶², and that orchestrates a complex catabolic response that increases mitochondrial biogenesis, enhances antioxidant defence and improves fatty-acid oxidation.

Mitochondrial oxidative stress and ageing

Mitochondrial networks are vulnerable to oxidative stress, resulting in impaired cellular function and increasing the chance of cell death. During ageing, mitochondrial electron transport chain (ETC) efficiency decreases, reducing ATP generation through increased proton leakage. Higher metabolic intensities correlated with higher proton conductance across the inner membrane of skeletal muscle mitochondria were found in long-lived mice, supporting the 'uncoupling to survive' theory¹¹.

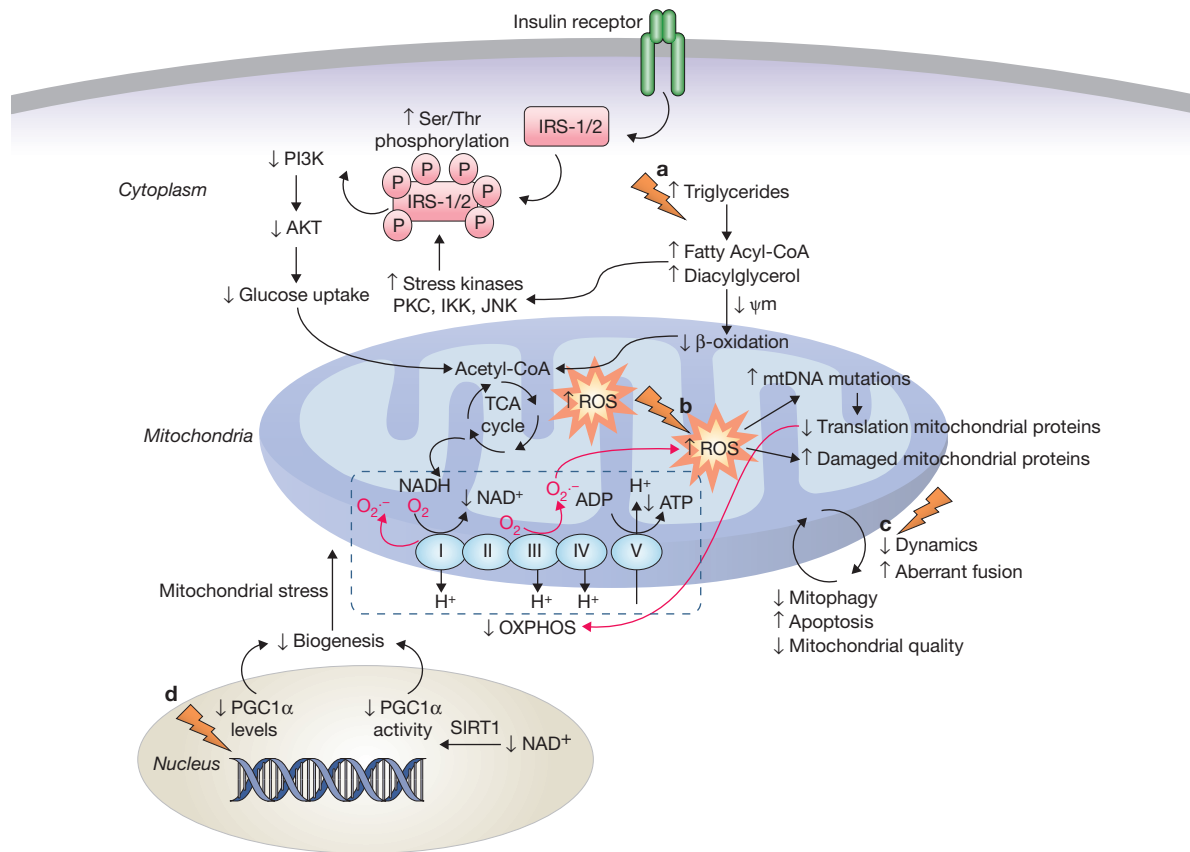


Figure 2 Effect of ageing on mitochondrial function. Multiple sources of damage (orange bolts, **a–d**) cause senescent mitochondrial phenotypes. **(a)** Nutrient surplus promotes the accumulation of lipid metabolite intermediates that overload and hyperpolarize the mitochondria. Incompletely oxidized lipid products from the tricarboxylic acid cycle (TCA) and reactive oxygen species (ROS) then build up, which causes oxidative stress to the respiratory system (OXPHOS; simplified electron transport chain shown for illustrative purposes). These intermediates also activate stress-sensitive kinases (protein kinase C, PKC; c-Jun N-terminal kinase, JNK; and IκB kinase, IKK), which

phosphorylate insulin receptor substrates (IRS-1/2). This disrupts the insulin signalling pathway, and glucose uptake and glycolysis in response to insulin. **(b)** ROS accumulation causes mutations to the mitochondrial DNA (mtDNA) pool, affecting mitochondrial translation and damage to proteins vital to OXPHOS and mitochondrial dynamics. **(c)** Impaired dynamics and decreased mitophagy with age leads to aberrant mitochondria. **(d)** Mitochondrial biogenesis, controlled by PGC1α, declines with age. Insulin resistance contributes to the development of a vicious cycle of a negative feedback loop by impairing biogenesis in addition to the TCA cycle and OXPHOS.

However, proton leakage in the ETC also diminishes the amount of reactive oxygen species (ROS) produced, with ROS being by-products of mitochondrial ATP generation resulting from oxygen reduction. Thus, reduction of ROS levels seems to be beneficial by dampening oxidative damage to mitochondria. The recruitment of uncoupling proteins (UCPs) relieves mitochondrial ROS by uncoupling the proton gradient. In particular, UCP1, found in metabolically active tissues such as brown fat, is activated by fatty acids and dissipates the proton gradient as heat, with a low rate of ATP production allowing fast substrate oxidation and thermogenesis⁶³. The production of superoxide (O_2^-) — a toxic by-product of respiration generated mostly by complexes I and III of the ETC — may also induce the uncoupling of mitochondria by activating UCP1 (or other UCPs) under certain conditions requiring fatty acids and inhibited by purine nucleotides⁶⁴. Accordingly, increasing energy expenditure by UCP1 overexpression in mouse muscle has beneficial effects on metabolic health, extending median but not maximal lifespan, decreasing adiposity, increasing both core temperature and metabolic rate, and lowering lymphoma incidence⁶⁵.

The ‘free radical’ theory of ageing proposes a causative role for ROS in accumulated cellular damage over time, and in age-associated

pathologies and functional decline of tissues. Supporting this theory, decreased ROS production from mitochondria has been observed in response to dietary restriction and linked to improved mitochondrial function in rodents. Despite the initial claim that increased respiration following dietary restriction is achieved by higher mitochondrial biogenesis⁶⁶, dietary restriction is now thought to preserve mitochondrial function by maintaining the integrity of existing cellular components and increasing the activity of oxidant scavengers such as catalase⁶⁷. Mice expressing human catalase in mitochondria are protected from oxidative damage, cardiac dysfunction and cataract development, and exhibit extended longevity⁶⁹. However, a conflicting study reports that overexpression of human catalase in all tissues is insufficient to extend lifespan⁷⁰. PGC1α also minimizes the build-up of ROS through the transcriptional regulation of numerous ROS-detoxifying enzymes⁶⁸. Other genetic models challenge the theory that a linear relationship exists between decreasing ROS and lifespan extension. Increasing mitochondrial ROS and oxidative damage do not cause progeria in mice^{71,72}, and multiple mouse models of increased antioxidant defences also do not extend lifespan⁷⁰. Furthermore, mice expressing a defective mitochondrial polymerase develop mitochondrial DNA (mtDNA) mutations

that accelerate ageing by impairing mitochondrial function without increasing ROS^{73–75}. Although the mtDNA mutator mouse studies initially challenged the free radical theory of ageing, recent conflicting evidence suggests that oxidative damage actually occurs in skeletal muscle of these mice⁷⁶. Following these controversies, the role of ROS in ageing is being revisited, mainly supported by evidence obtained in *Caenorhabditis elegans* suggesting that high ROS levels may act as an important signal that triggers a gene expression pattern promoting survival under stressful conditions⁷⁷.

Activation of the mitochondrial UPR and longevity

Long-lived mouse models provide supporting evidence for the role of improving mitochondrial biogenesis and function in forestalling ageing. Paradoxically, in *C. elegans*, lifespan can be increased by reducing mitochondrial respiration through reduced function of nuclear genes encoding ETC components^{78,79}. In particular, neuronal knockdown of COX activates the mitochondrial unfolded protein response (UPR^{mt}), a mitochondrial proteostasis mechanism that confers increased longevity to worms^{80,81}. This pathway is part of the mitochondrial quality control machinery that detects misfolded proteins in the mitochondria, initiating a signalling cascade that leads to the transcription of protective genes⁸². Notably, 13 subunits of the oxidative phosphorylation complex are encoded by mtDNA, whereas the remaining mitochondrial proteins (of which there are more than 1,000) are encoded by nuclear DNA and imported into mitochondria to achieve proper folding. The UPR^{mt} senses imbalances in the stoichiometry of proteins in the mitochondrial compartment and relieves the stress by retrograde signalling to the nucleus through the degradation of misfolded proteins into peptides by the ClpP1 protease, consequently promoting transcriptional activation of nuclear-encoded chaperones residing in the mitochondrial matrix, such as heat shock proteins 60 (HSP60) and 70 (HSP70). This surveillance pathway is conserved in mammalian cells⁸³, but whether it plays a pro-longevity role in mammals is unknown. In support of its possible role in mouse lifespan regulation, low expression of a mouse mitochondrial ribosomal protein (Mrps5) triggers the UPR^{mt} and correlates with increased lifespan of inbred progenies of a cross between C57BL/6J and DBA/2J mice⁸⁴.

Evidence from mouse longevity studies suggests that impairing mitochondrial function can improve lifespan; however, it is unknown whether the UPR^{mt} is causal for the longevity increase. Constitutive ablation of SURF1 (surfeit 1), a putative assembly factor specific to COX, results in mild deficiency of COX in mice and significantly extended mouse longevity⁸⁵. This is accompanied by a strong depletion of PGC1 α levels, mtDNA content, citrate synthase activity and oxidative-phosphorylation-related genes in these mice, indicative of a decrease in respiration and mitochondrial biogenesis⁵⁸. Reduction of COQ7 (coenzyme Q7, known as Mcl1 in mice), an enzyme responsible for the biosynthesis of ubiquinone (an ETC component), also extends mouse lifespan concomitant with ETC dysfunction and decreased mitochondrial coupling of respiration and ATP synthesis⁸⁶. Although these studies confirm that decreased respiration has beneficial effects on mouse longevity, it is unknown whether the UPR^{mt} following mitochondrial dysfunction is required for longevity.

Strikingly, levels of NAD⁺ were also decreased in the Mcl1^{+/-} mice. It is well established that NAD⁺ plays crucial roles in mediating mitochondrial energy metabolism by donating electrons to the ETC or by

acting as a coenzyme for rate-limiting citric acid (TCA) cycle enzymes. Increasing NAD⁺ levels, by supplementation of a NAD⁺ precursor or through a drug such as resveratrol, improves mitochondrial function by activating SIRT1 in mice, and protects against the metabolic damage of high-fat feeding^{87–90}. Thus, high NAD⁺ levels correspond to mitochondrial biogenesis and metabolic flexibility, ultimately improving healthspan. Paradoxically, Mcl1^{+/-} mice are long-lived despite their depleted NAD⁺ levels. The mechanisms underlying this lifespan extension are unclear, and it is plausible that a different longevity paradigm independent of mitochondrial biogenesis, such as UPR^{mt}, is required to promote lifespan of these mice. The relationship between UPR^{mt} and mitochondrial biogenesis is, however, still under debate. Rapamycin and resveratrol treatment, which extend lifespan and metabolic fitness, increased respiration in mammalian cells and induced the UPR^{mt} (ref. 84). Thus, it is unclear whether UPR^{mt} in mammals predicts longevity and can be triggered as a cell survival mechanism following severe mitochondrial impairment, or as a response to mitonuclear imbalance that facilitates biogenesis. More investigation is necessary to elucidate the physiological implication of the UPR^{mt} in mammalian ageing.

Insulin resistance, mitochondrial perturbations and dynamics

Mitochondrial dysfunction has been implicated strongly in the onset of metabolic diseases. Cells cope with nutrient supply by increasing mitochondrial content, but persistent nutrient surplus can overload the mitochondria and cause dysfunction, a phenomenon also observed with ageing, as mitochondrial capacity declines (Fig. 2). This functional decline coincides with a global reduction in oxidative capacity of the skeletal muscle of patients with type 2 diabetes⁹¹, and contributes to insulin resistance⁵¹. Energy excess overloads and hyperpolarizes mitochondria, leading to the accumulation of fatty-acyl-CoA and diacylglycerol, causing excessive production of ROS. These mitochondrial metabolites activate stress-sensing-kinases such as c-Jun N-terminal kinase and protein kinase C in the cytoplasm (which phosphorylate insulin receptor substrates), impairing insulin signalling in glucose-consuming tissues such as skeletal muscle and liver^{92,93}. Insulin-stimulated glucose transport is inhibited in these tissues and reduces peripheral energy metabolism.

Furthermore, decreased brain insulin signalling is also observed in ageing and contributes to neurodegenerative disorders, as a consequence of insulin resistance and decreased insulin transport through the blood–brain barrier⁹⁴. Abnormal insulin metabolism in the brain might originate from mitochondrial dysfunction, as the brain is highly vulnerable to oxidative stress and elevated ROS levels can aggravate insulin resistance^{95,96}. Similar observations were made in the hypothalamus and hippocampus of mice fed a high-fat diet^{97,98}. Downregulation of the mitochondrial chaperone protein HSP60 by excessive adipokine stimuli plays a central role in the onset of oxidative stress leading to mitochondrial dysfunction and hypothalamic insulin resistance⁹⁹. Whether chaperone levels are also regulated by age-associated stress in the brain and confer mitochondrial dysfunction and deficits in brain energy metabolism remains undetermined.

Mitochondrial architecture is tightly regulated by the dynamic opposing processes of fusion and fission. Mitochondrial dynamics reflect bioenergetic adaptation to metabolic demand, facilitating mixing and exchange of mitochondrial contents such as mtDNA or metabolites¹⁰⁰. Mitochondrial morphology and density are modulated

by variations in nutrient availability, with starvation triggering a fused state that facilitates autophagy¹⁰¹. These changes in mitochondrial architecture can affect oxidative phosphorylation complex assembly and ATP synthesis, allowing mitochondria to reorganize and dispose of damaged elements through mitophagy, an autophagic process that eliminates damaged mitochondria¹⁰². Mitophagy prevents cell apoptosis or necrosis following excessive ROS accumulation, pro-inflammatory signals or permeabilization of mitochondrial membranes, but the efficiency of autophagy declines with age¹⁰³. Fission allows the segregation of damaged mitochondria and their recycling through autophagic processes, thereby ensuring mitochondria turnover and cellular viability, whereas fusion counterbalances functional defects and allows genetic complementation. Oversized abnormal mitochondria are present in senescent cells, as a result of dysfunctional mitochondrial degradation¹⁰⁴. Youthful mitochondrial fragmentation can be restored in senescent cells by depleting critical mediators of fusion, the dynamin-related proteins FIS1 (fission 1 (mitochondrial outer membrane) homolog) and OPA1 (optic atrophy 1). Recently, lack of the pro-fusion mitofusins Mfn1 and Mfn2 has been implicated in regulation of whole-body energy homeostasis in distinct populations of hypothalamic neurons in mice^{105–107}. Mfn2 deficiency prevents changes in mitochondrial dynamics and communication with the endoplasmic reticulum (ER), causing ER stress, insulin resistance in skeletal muscle and liver tissues, and hypothalamic leptin resistance, thus promoting obesity^{105–109}. Mitochondrial health relies on a tight contact with the ER that provides critical resources to the mitochondria such as mitochondrial lipids and calcium ions, which are involved in mitochondrial dynamics and regulation of the TCA cycle. Although gradual impairment of mitochondrial dynamics with age is a proposed hypothesis for age-associated mitochondrial dysfunction, additional layers of regulation may exist, including ER-mitochondria tethering and impaired insulin signalling due to ER stress, resulting in cell-autonomous inflammation (as observed in type 2 diabetes)^{110,111}. One of the main pro-inflammatory transcriptional programs induced following ER stress is the NF- κ B (nuclear factor κ B) signalling pathway¹¹². Interestingly, restoring hypothalamic immunity upon ageing through the blockade of NF- κ B signalling in microglia is sufficient to extend lifespan in mice¹¹³. It will be critical to define whether metabolic decline during ageing is mediated by the disruption of intricate connections between mitochondria and the ER, and the onset of ER stress and inflammation.

Conclusions and future perspectives

In light of the evidence reviewed here, progressive mitochondrial dysfunction is implicated in the deterioration of insulin sensitivity during normal ageing, consequently impairing insulin signalling in central and peripheral tissues, and contributing to a loss of metabolic flexibility. Thus, restoring mitochondrial biogenesis is likely to provide a therapeutic avenue to rejuvenate metabolism. However, in conflicting studies, reduced respiration and biogenesis throughout life can also increase lifespan, indicating an additional level of complexity. Epigenetic regulation is now emerging as a potent factor influencing mitochondrial biogenesis or integrity following the lack of exercise and overfeeding. Notably, DNA hypermethylation suppresses the expression of Pgc1 α and Tfam in mice, and microRNAs have the ability to silence gene expression of Ucp2 and Mfn2, thus negatively affecting

biogenesis, oxidative phosphorylation and mitochondrial dynamics¹¹⁴. Whether such epigenetic signatures are characteristic of the ageing process remains poorly understood; thus, it will be essential to assess the chain of events that leads to impaired transcriptional networks that maintain biogenesis (nuclear and mitochondrial transcription) as well as mitochondrial quality control machinery (dynamics, mitophagy and the UPR^{mt}). In particular, we are beginning to envision that these processes are likely to co-regulate each other, as highlighted by the reliance on fusion and fission events in order to remove damaged mitochondria through mitophagy and the dependence of biogenesis on protein translocation resulting from mitochondrial dynamics. These preliminary discoveries are identifying novel therapeutic perspectives in the effort to delay the onset of metabolic decline and age-associated metabolic diseases.

ACKNOWLEDGMENTS

We apologize to our colleagues for omitting numerous references due to space limitations. We thank Carsten Merkwirth and Kristen Berendzen for their helpful comments on the manuscript. C.E.R is supported by the American Diabetes Association Pathway to Stop Diabetes Grant 1-15-INI-12.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

- López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M. & Kroemer, G. The hallmarks of aging. *Cell* **153**, 1194–1217 (2013).
- De Cabo, R., Carmona-Gutierrez, D., Bernier, M., Hall, M. N. & Madeo, F. The search for antiaging interventions: from elixirs to fasting regimens. *Cell* **157**, 1515–1526 (2014).
- Selman, C. & Withers, D. J. Mammalian models of extended healthy lifespan. *Philos. Trans. R. Soc. Lond. Ser. B. Biol. Sci.* **366**, 99–107 (2011).
- Rowe, J. W., Minaker, K. L., Pallotta, J. A. & Flier, J. S. Characterization of the insulin resistance of aging. *J. Clin. Invest.* **71**, 1581–1587 (1983).
- Fink, R. I., Kolterman, O. G., Griffin, J. & Olefsky, J. M. Mechanisms of insulin resistance in aging. *J. Clin. Invest.* **71**, 1523–1535 (1983).
- Houtkooper, R. H. *et al.* The metabolic footprint of aging in mice. *Sci. Rep.* **1**, 134 (2011).
- Riera, C. E. *et al.* TRPV1 pain receptors regulate longevity and metabolism by neuropeptide signaling. *Cell* **157**, 1023–1036 (2014).
- Kurosu, H. *et al.* Suppression of aging in mice by the hormone klotho. *Science* **309**, 1829–1833 (2005).
- Selman, C. *et al.* Evidence for lifespan extension and delayed age-related biomarkers in insulin receptor substrate 1 null mice. *FASEB J.* **22**, 807–818 (2008).
- Taguchi, A., Wartschow, L. M. & White, M. F. Brain IRS2 signaling coordinates life span and nutrient homeostasis. *Science* **317**, 369–372 (2007).
- Speakman, J. R. *et al.* Uncoupled and surviving: individual mice with high metabolism have greater mitochondrial uncoupling and live longer. *Aging Cell* **3**, 87–95 (2004).
- Galgani, J. E., Moro, C. & Ravussin, E. Metabolic flexibility and insulin resistance. *Am. J. Physiol. Endocrinol. Metab.* **295**, 1009–1017 (2008).
- Kondratov, R. V., Kondratova, A. A., Gorbacheva, V. Y., Vykhovanets, O. V. & Antoch, M. P. Early aging and age-related pathologies in mice deficient in BMAL1, the core component of the circadian clock. *Genes Dev.* **20**, 1868–1873 (2006).
- Chang, H.-C. & Guarente, L. SIRT1 mediates central circadian control in the SCN by a mechanism that decays with aging. *Cell* **153**, 1448–1460 (2013).
- Sahar, S. & Sassone-Corsi, P. Metabolism and cancer: the circadian clock connection. *Nat. Rev. Cancer* **9**, 886–896 (2009).
- Fontana, L., Partridge, L. & Longo, V. D. Dietary restriction, growth factors and aging: from yeast to humans. *Science* **328**, 321–326 (2010).
- Mair, W. & Dillin, A. Aging and survival: the genetics of life span extension by dietary restriction. *Annu. Rev. Biochem.* **77**, 727–754 (2008).
- Bruss, M. D., Khambatta, C. F., Ruby, M. A., Aggarwal, I. & Hellerstein, M. K. Calorie restriction increases fatty acid synthesis and whole body fat oxidation rates. *Am. J. Physiol. Endocrinol. Metab.* **298**, E108–E116 (2010).
- Blüher, M. Extended longevity in mice lacking the insulin receptor in adipose tissue. *Science* **299**, 572–574 (2003).
- Foukas, L. C. *et al.* Long-term p110 β PI3K inactivation exerts a beneficial effect on metabolism. *EMBO Mol. Med.* **5**, 563–571 (2013).
- Zhang, Y. *et al.* The starvation hormone, fibroblast growth factor-21, extends lifespan in mice. *eLife* **1**, e00065 (2012).
- Yuan, R. *et al.* Aging in inbred strains of mice: study design and interim report on median lifespans and circulating IGF1 levels. *Aging Cell* **8**, 277–287 (2009).
- Berryman, D. E., Christiansen, J. S., Johannsson, G., Thorne, M. O. & Kopchick, J. J. Role of the GH/IGF-1 axis in lifespan and healthspan: lessons from animal models. *Growth Horm. IGF Res.* **18**, 455–471 (2008).
- Hsieh, C.-C., DeFord, J. H., Flurkey, K., Harrison, D. E. & Papaconstantinou, J. Implications for the insulin signaling pathway in Snell dwarf mouse longevity: a similarity with the *C. elegans* longevity paradigm. *Mech. Ageing Dev.* **123**, 1229–1244 (2002).

25. Dominici, F. P., Hauck, S., Argentino, D. P., Bartke, A. & Turyn, D. Increased insulin sensitivity and upregulation of insulin receptor, insulin receptor substrate (IRS)-1 and IRS-2 in liver of Ames dwarf mice. *J. Endocrinol.* **173**, 81–94 (2002).
26. Shah, J. H. & Cerchio, G. M. Hypoinsulinemia of hypothyroidism. *Arch. Intern. Med.* **132**, 657–661 (1973).
27. Tsugane, S. & Inoue, M. Insulin resistance and cancer: epidemiological evidence. *Cancer Sci.* **101**, 1073–1079 (2010).
28. Naderali, E. K., Ratcliffe, S. H. & Dale, M. C. Obesity and Alzheimer's disease: a link between body weight and cognitive function in old age. *Am. J. Alzheimers Dis. Other Demen.* **24**, 445–449 (2009).
29. Ortega-Molina, A. *et al.* Pten positively regulates brown adipose function, energy expenditure, and longevity. *Cell Metab.* **15**, 382–394 (2012).
30. Coschigano, K. T., Clemmons, D., Bellush, L. L. & Kopchick, J. J. Assessment of growth parameters and life span of GHR/BP gene-disrupted mice. *Endocrinology* **141**, 2608–2613 (2000).
31. Ikeno, Y., Bronson, R. T., Hubbard, G. B., Lee, S. & Bartke, A. Delayed occurrence of fatal neoplastic diseases in ames dwarf mice: correlation to extended longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **58**, 291–296 (2003).
32. Ikeno, Y. *et al.* Reduced incidence and delayed occurrence of fatal neoplastic diseases in growth hormone receptor/binding protein knockout mice. *J. Gerontol. A Biol. Sci. Med. Sci.* **64**, 522–529 (2009).
33. Harrison, D. E. *et al.* Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* **460**, 392–395 (2009).
34. Miller, R. A. *et al.* Rapamycin, but not resveratrol or simvastatin, extends life span of genetically heterogeneous mice. *J. Gerontol. A Biol. Sci. Med. Sci.* **66A**, 191–201 (2011).
35. Corradetti, M. N. & Guan, K.-L. Upstream of the mammalian target of rapamycin: do all roads pass through mTOR? *Oncogene* **25**, 6347–6360 (2006).
36. Lamming, D. W. *et al.* Rapamycin-induced insulin resistance is mediated by mTORC2 loss and uncoupled from longevity. *Science* **335**, 1638–1643 (2012).
37. Roy, J., Paquette, J.-S., Fortin, J.-F. & Tremblay, M. J. The immunosuppressant rapamycin represses human immunodeficiency virus type 1 replication. *Antimicrob. Agents Chemother.* **46**, 3447–3455 (2002).
38. Fang, Y. *et al.* Duration of rapamycin treatment has differential effects on metabolism in mice. *Cell Metab.* **17**, 456–462 (2013).
39. Pyo, J.-O. *et al.* Overexpression of Atg5 in mice activates autophagy and extends lifespan. *Nat. Commun.* **4**, 2300 (2013).
40. Selman, C. *et al.* Ribosomal protein S6 kinase 1 signaling regulates mammalian life span. *Science* **326**, 140–144 (2009).
41. Wu, J. J. *et al.* Increased mammalian lifespan and a segmental and tissue-specific slowing of aging after genetic reduction of mTOR expression. *Cell Rep.* **4**, 913–920 (2013).
42. Saltiel, A. R. & Kahn, C. R. Insulin signalling and the regulation of glucose and lipid metabolism. *Nature* **414**, 799–806 (2001).
43. Bratic, A. & Larsson, N.-G. The role of mitochondria in aging. *J. Clin. Invest.* **123**, 951–957 (2013).
44. Schapira, A. H. V. Mitochondrial diseases. *Lancet* **379**, 1825–1834 (2012).
45. Herbenner, G. H. A morphometric study of age-dependent changes in mitochondrial population of mouse liver and heart. *J. Gerontol.* **31**, 8–12 (1976).
46. Lanza, I. R. & Nair, K. S. Mitochondrial function as a determinant of life span. *Pflüg. Arch. Eur. J. Physiol.* **459**, 277–289 (2010).
47. Conley, K. E., Jubrias, S. A. & Esselman, P. C. Oxidative capacity and ageing in human muscle. *J. Physiol.* **526**, 203–210 (2000).
48. Lin, J. *et al.* Transcriptional co-activator PGC-1 α drives the formation of slow-twitch muscle fibres. *Nature* **418**, 797–801 (2002).
49. Puigserver, P. *et al.* A cold-inducible coactivator of nuclear receptors linked to adaptive thermogenesis. *Cell* **92**, 829–839 (1998).
50. Mootha, V. K. *et al.* PGC-1 α -responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat. Genet.* **34**, 267–273 (2003).
51. Patti, M. E. *et al.* Coordinated reduction of genes of oxidative metabolism in humans with insulin resistance and diabetes: potential role of PGC1 and NRF1. *Proc. Natl Acad. Sci. USA* **100**, 8466–8471 (2003).
52. Wenz, T., Rossi, S. G., Rotundo, R. L., Spiegelman, B. M. & Moraes, C. T. Increased muscle PGC-1 α expression protects from sarcopenia and metabolic disease during aging. *Proc. Natl Acad. Sci. USA* **106**, 20405–20410 (2009).
53. Anderson, R. & Prolla, T. PGC-1 α in aging and anti-aging interventions. *Biochim. Biophys. Acta* **1790**, 1059–1066 (2009).
54. Austin, S. & St-Pierre, J. PGC1 α and mitochondrial metabolism — emerging concepts and relevance in ageing and neurodegenerative disorders. *J. Cell Sci.* **125**, 4963–4971 (2012).
55. Cantó, C. *et al.* AMPK regulates energy expenditure by modulating NAD⁺ metabolism and SIRT1 activity. *Nature* **458**, 1056–1060 (2009).
56. Jäger, S., Handschin, C., St-Pierre, J. & Spiegelman, B. M. AMP-activated protein kinase (AMPK) action in skeletal muscle via direct phosphorylation of PGC-1 α . *Proc. Natl Acad. Sci. USA* **104**, 12017–12022 (2007).
57. Narkar, V. A. *et al.* AMPK and PPAR δ agonists are exercise mimetics. *Cell* **134**, 405–415 (2008).
58. Viscomi, C. *et al.* In vivo correction of COX deficiency by activation of the AMPK/PGC-1 α axis. *Cell Metab.* **14**, 80–90 (2011).
59. Martin-Montalvo, A. *et al.* Metformin improves healthspan and lifespan in mice. *Nat. Commun.* **4**, 2192 (2013).
60. Kalender, A. *et al.* Metformin, independent of AMPK, inhibits mTORC1 in a rag GTPase-dependent manner. *Cell Metab.* **11**, 390–401 (2010).
61. Cantó, C. *et al.* Interdependence of AMPK and SIRT1 for metabolic adaptation to fasting and exercise in skeletal muscle. *Cell Metab.* **11**, 213–219 (2010).
62. Price, N. L. *et al.* SIRT1 is required for AMPK activation and the beneficial effects of resveratrol on mitochondrial function. *Cell Metab.* **15**, 675–690 (2012).
63. Nicholls, D. G., Bernson, V. S. M. & Heaton, G. M. in *Effectors of Thermogenesis* (eds. Girardier, L. & Seydoux, J.) 89–93 (Birkhäuser Basel, 1978).
64. Echtay, K. S. *et al.* Superoxide activates mitochondrial uncoupling proteins. *Nature* **415**, 96–99 (2002).
65. Gates, A. C. *et al.* Respiratory uncoupling in skeletal muscle delays death and diminishes age-related disease. *Cell Metab.* **6**, 497–505 (2007).
66. Nisoli, E. *et al.* Calorie restriction promotes mitochondrial biogenesis by inducing the expression of eNOS. *Science* **310**, 314–317 (2005).
67. Lanza, I. R. *et al.* Chronic caloric restriction preserves mitochondrial function in senescence without increasing mitochondrial biogenesis. *Cell Metab.* **16**, 777–788 (2012).
68. St-Pierre, J. *et al.* Bioenergetic analysis of peroxisome proliferator-activated receptor γ coactivators 1 α and 1 β (PGC-1 α and PGC-1 β) in muscle cells. *J. Biol. Chem.* **278**, 26597–26603 (2003).
69. Schriener, S. E. *et al.* Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* **308**, 1909–1911 (2005).
70. Pérez, V. I. *et al.* The overexpression of major antioxidant enzymes does not extend the lifespan of mice. *Aging Cell* **8**, 73–75 (2009).
71. Van Remmen, H. *et al.* Life-long reduction in MnSOD activity results in increased DNA damage and higher incidence of cancer but does not accelerate aging. *Physiol. Genomics* **16**, 29–37 (2003).
72. Zhang, Y. *et al.* Mice deficient in both Mn superoxide dismutase and glutathione peroxidase-1 have increased oxidative damage and a greater incidence of pathology but no reduction in longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **64**, 1212–1220 (2009).
73. Edgar, D. *et al.* Random point mutations with major effects on protein-coding genes are the driving force behind premature aging in mtDNA mutator mice. *Cell Metab.* **10**, 131–138 (2009).
74. Trifunovic, A. *et al.* Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature* **429**, 417–423 (2004).
75. Vermulst, M. *et al.* DNA deletions and clonal mutations drive premature aging in mitochondrial mutator mice. *Nat. Genet.* **40**, 392–394 (2008).
76. Kolesar, J. E. *et al.* Defects in mitochondrial DNA replication and oxidative damage in muscle of mtDNA mutator mice. *Free Radic. Biol. Med.* **75**, 241–251 (2014).
77. Yee, C., Yang, W. & Hekimi, S. The intrinsic apoptosis pathway mediates the pro-longevity response to mitochondrial ROS in *C. elegans*. *Cell* **157**, 897–909 (2014).
78. Dillin, A. *et al.* Rates of behavior and aging specified by mitochondrial function during development. *Science* **298**, 2398–2401 (2002).
79. Lee, S. S. *et al.* A systematic RNAi screen identifies a critical role for mitochondria in *C. elegans* longevity. *Nat. Genet.* **33**, 40–48 (2003).
80. Durieux, J., Wolff, S. & Dillin, A. The cell-non-autonomous nature of electron transport chain-mediated longevity. *Cell* **144**, 79–91 (2011).
81. Yoneda, T. *et al.* Compartment-specific perturbation of protein handling activates genes encoding mitochondrial chaperones. *J. Cell Sci.* **117**, 4055–4066 (2004).
82. Mottis, A., Jovaisaite, V. & Auwerx, J. The mitochondrial unfolded protein response in mammalian physiology. *Mamm. Genome* **25**, 424–433 (2014).
83. Zhao, Q. *et al.* A mitochondrial specific stress response in mammalian cells. *EMBO J.* **21**, 4411–4419 (2002).
84. Houtkooper, R. H. *et al.* Mitonuclear protein imbalance as a conserved longevity mechanism. *Nature* **497**, 451–457 (2013).
85. Dell'agnello, C. *et al.* Increased longevity and refractoriness to Ca²⁺-dependent neurodegeneration in Surf1 knockout mice. *Hum. Mol. Genet.* **16**, 431–444 (2007).
86. Lapointe, J. & Hekimi, S. Early mitochondrial dysfunction in long-lived Mcl1^{+/+} mice. *J. Biol. Chem.* **283**, 26217–26227 (2008).
87. Bai, P. *et al.* PARP-2 regulates SIRT1 expression and whole-body energy expenditure. *Cell Metab.* **13**, 450–460 (2011).
88. Bai, P. *et al.* PARP-1 inhibition increases mitochondrial metabolism through SIRT1 activation. *Cell Metab.* **13**, 461–468 (2011).
89. Lagouge, M. *et al.* Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1 α . *Cell* **127**, 1109–1122 (2006).
90. Yoshino, J., Mills, K. F., Yoon, M. J. & Imai, S. Nicotinamide mononucleotide, a key NAD⁺ intermediate, treats the pathophysiology of diet- and age-induced diabetes in mice. *Cell Metab.* **14**, 528–536 (2011).
91. Kelley, D. E., He, J., Menshikova, E. V. & Ritov, V. B. Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. *Diabetes* **51**, 2944–2950 (2002).
92. Fisher-Wellman, K. H. & Neuffer, P. D. Linking mitochondrial bioenergetics to insulin resistance via redox biology. *Trends Endocrinol. Metab.* **23**, 142–153 (2012).
93. Lowell, B. B. & Shulman, G. I. Mitochondrial dysfunction and type 2 diabetes. *Science* **307**, 384–387 (2005).
94. Cholerton, B., Baker, L. D. & Craft, S. Insulin resistance and pathological brain aging. *Diabet. Med.* **28**, 1463–1475 (2011).
95. Evans, J. L., Maddux, B. A. & Goldfine, I. D. The molecular basis for oxidative stress-induced insulin resistance. *Antioxid. Redox Signal.* **7**, 1040–1052 (2005).
96. Halliwell, B. Reactive oxygen species and the central nervous system. *J. Neurochem.* **59**, 1609–1623 (1992).
97. Pipatpiboon, N., Pratchayasakul, W., Chattipakorn, N. & Chattipakorn, S. C. PPAR γ agonist improves neuronal insulin receptor function in hippocampus and brain mitochondria function in rats with insulin resistance induced by long term high-fat diets. *Endocrinology* **153**, 329–338 (2012).

98. Thaler, J. P. *et al.* Obesity is associated with hypothalamic injury in rodents and humans. *J. Clin. Invest.* **122**, 153–162 (2012).
99. Kleinridders, A. *et al.* Leptin regulation of *Hsp60* impacts hypothalamic insulin signaling. *J. Clin. Invest.* **123**, 4667–4680 (2013).
100. Chan, D. C. Fusion and fission: interlinked processes critical for mitochondrial health. *Annu. Rev. Genet.* **46**, 265–287 (2012).
101. Gomes, L. C., Di Benedetto, G. & Scorrano, L. During autophagy mitochondria elongate, are spared from degradation and sustain cell viability. *Nat. Cell Biol.* **13**, 589–598 (2011).
102. Green, D. R., Galluzzi, L. & Kroemer, G. Mitochondria and the autophagy-inflammation-cell death axis in organismal aging. *Science* **333**, 1109–1112 (2011).
103. Zhang, J. Autophagy and mitophagy in cellular damage control. *Redox Biol.* **1**, 19–23 (2013).
104. Lee, S. *et al.* Mitochondrial fission and fusion mediators, hFis1 and OPA1, modulate cellular senescence. *J. Biol. Chem.* **282**, 22977–22983 (2007).
105. Dietrich, M. O., Liu, Z-W. & Horvath, T. L. Mitochondrial dynamics controlled by mitofusins regulate *Agrp* neuronal activity and diet-induced obesity. *Cell* **155**, 188–199 (2013).
106. Schneeberger, M. *et al.* Mitofusin 2 in POMC neurons connects ER stress with leptin resistance and energy imbalance. *Cell* **155**, 172–187 (2013).
107. De Brito, O. M. & Scorrano, L. Mitofusin 2 tethers endoplasmic reticulum to mitochondria. *Nature* **456**, 605–610 (2008).
108. Sebastián, D. *et al.* Mitofusin 2 (*Mfn2*) links mitochondrial and endoplasmic reticulum function with insulin signaling and is essential for normal glucose homeostasis. *Proc. Natl Acad. Sci. USA* **109**, 5523–5528 (2012).
109. Muñoz, J. P. *et al.* *Mfn2* modulates the UPR and mitochondrial function via repression of PERK. *EMBO J.* **32**, 2348–2361 (2013).
110. Ozcan, U. *et al.* Endoplasmic reticulum stress links obesity, insulin action, and type 2 diabetes. *Science* **306**, 457–461 (2004).
111. Garg, A. D. *et al.* ER stress-induced inflammation: does it aid or impede disease progression? *Trends Mol. Med.* **18**, 589–598 (2012).
112. Zhang, K. & Kaufman, R. J. From endoplasmic-reticulum stress to the inflammatory response. *Nature* **454**, 455–462 (2008).
113. Zhang, G. *et al.* Hypothalamic programming of systemic ageing involving IKK- β , NF- κ B and GnRH. *Nature* **497**, 211–216 (2013).
114. Cheng, Z. & Almeida, F. A. Mitochondrial alteration in type 2 diabetes and obesity: an epigenetic link. *Cell Cycle* **13**, 890–897 (2014).